

*Original Research***The Effect of Supplementation of Oregano Oil and Probiotic on Intestinal Microbes (Namely *E. coli* spp., *Salmonella* spp., *Clostridia* spp.) of the Broiler Chicken****Harshal N. Rewatkar*, S. M. Wankhede, J. L. Agashe¹, R. M. Padole², A. D. Jadhao and G. M. Jadhao**

Department of Animal Nutrition, Post Graduate Institute of Veterinary and Animal Sciences, Akola, Maharashtra Animal and Fishery Sciences University, Nagpur, Maharashtra, INDIA

¹Department of Poultry Science²Department of Pathology***Corresponding author:** rewatkarharshal@gmail.com

Rec. Date:	May 04, 2019 06:36
Accept Date:	Jun 06, 2019 11:06
DOI	10.5455/ijlr.20190504063600

Abstract

To evaluate the effect of oregano oil and probiotic on total microbial count of intestine namely *E. coli*, *Salmonella* and *Clostridia*. On 42nd day, 8 birds from each treatment were slaughtered by decapitation. The digestive tracts were dissected and described Caecal content samples were taken aseptically and were transferred into sterile plastic bags and immediately transported in cold chain to the laboratory, samples diluted, cultured and checked for the presence of organisms in samples (*E. coli*, *Salmonella*, *Clostridia*) and compared with the control after incubation. Results were expressed as log₁₀ colony-forming units per gram of ileal digesta (log₁₀ CFU/gram). The intestinal microbial count namely *E. coli*, *Salmonella*, *Clostridia* were significantly decreased in T₃ than in groups T₂, T₁ and control. It was concluded that feeding of Oregano essential oil and probiotic- *Saccharomyces cerevisiae* exhibited beneficial effect on GIT and decreased the harmful microbial count of the intestine in the broiler chicken.

Key words: Broiler, Intestinal Microbes, Oregano Essential Oil, Probiotic**How to cite:** Rewatkar, H., Wankhede, S., Agashe, J., Padole, R., Jadhao, A., & Jadhao, G. (2019). The Effect of Supplementation of Oregano Oil and Probiotic on Intestinal Microbes (*E. coli* spp., *Salmonella* spp., *Clostridia* spp.) of the Broiler Chicken. International Journal of Livestock Research, 9(7), 77-84. Doi: 10.5455/ijlr.20190504063600**Introduction**

The poultry industry demands maximum performance from broilers as well as layer. Now a days, essential oils (EOs) are used in poultry feed, these have antimicrobial, antioxidant, antifungal, antiparasitic and antiviral properties. Beside this, other beneficial effects of EOs include appetite stimulation, improvement of enzyme secretion related to food digestion and immune response activation. Recently, use of EOs in

broiler chickens has drawn attention due to their advantageous properties. Many kinds of natural substances, prebiotics and probiotics have been supplemented to broilers to increase poultry production by activating intestinal function (B. Shanmuga Priya* and S. Saravana Babu, 2013). Awaad *et al.* (2014) the effect of a specific combination (SC) of Carvacrol (active constitute of oregano essential oil), cinnamaldehyde on productive performance and immune response in broiler chickens. Edward *et al.* (2002) the effect of dietary supplementation of oregano oil (*Origanum sp.*) medicinal plant on, not significant effects on blood biochemical parameters and immune system of broiler chickens. Manafi *et al.* (2016) the commercial probiotic multispecies probiotic having four *Bacillus species* and *Saccharomyces boulardii* (Microguard®) with a (Protexin®) and a commonly used antibiotic in broilers probiotic having four *Bacillus species* and *Saccharomyces boulardii* (Microguard®) at 150 g/ton showed increase in blood serum concentration of high density lipoprotein, triglyceride in probiotic-supplemented broilers. The probiotics are used in poultry for “competitive/exclusion” of bacterial pathogens (Barrow P *et al.*, 1992). Probiotics are living microorganisms which upon ingestion in adequate amounts confer health benefits to host (FAO/WHO 2002). Exactly how supplemental dietary microbial products function in the digestive system is not known, but some suggested mechanisms are that they- 1) provide nutrients, 2) aid in digesting foods and 3) inhibit harmful bacteria (Owings *et al.*, 1990). It creates gut conditions that suppress harmful microorganisms and favor beneficial ones (Line *et al.*, 1998; Mead *et al.*, 2000). The antibacterial properties of EOs are well documented (Tihonen *et al.*, 2010). The supplementation of curd as a probiotics showed a positive effect on blood biochemical and caecal *Lactobacilli* and *E. coli* count in broiler chicken. (Chaudhary *et al.*, 2017), So, the present study is aimed at investigating whether oregano oil and probiotic could decrease in microbial count namely *E. coli spp.*, *Salmonella spp.*, *Clostridia spp.* of intestine of birds or not.

Materials and Method

Birds and Housing

The present study was conducted to evaluate the effect of feeding oregano (*Origanum vulgare*) oil as phytobiotic growth promoter with probiotic on intestinal histomorphology of Broiler Chicken. Two hundred forty straight straight run “Ven-Cobb 400Y” strain commercial day-old broiler chicks were equally and randomly distributed into four groups. Each treatment was subjected to four replicates with fifteen chicks.

Management of Bird

Prior to experimental trial, the experimental broiler shed, premises and the equipments were thoroughly cleaned and disinfected. Saw dust was provided as a bedding material for the chicks. Immediately after arrival, all the chicks were provided glucose through drinking water. The desired brooding temperature was maintained using incandescent bulbs. All the groups were provided similar environmental and management

conditions throughout the experimental period. The experimental chicks were offered ad-libitum feed and fresh water. The standard floor, feeding and watering space was provided throughout the experimental period depending on their age group. The experimental chicks were housed in 16 different pens. Each pen was accommodating 15 birds. The experimental birds were vaccinated against Ranikhet disease through intraocular route on 7th day with B1 strain, Infectious Bursal disease (IBD) vaccine on 14th day of age by intraocular route and booster vaccination of Infectious Bursal disease (IBD) was done on 21st day and vaccination of Ranikhet disease with lasota strain on 28th day through drinking water.

Procurement of Feed Ingredients

The good quality feed ingredients were procured from local market for preparation of experimental diets. Oregano essential oil was procured from Karma Essential Oil Pharmaceuticals. The chemical analysis of Maize and Soybean was carried out as per AOAC, 2012 (Table 2). The diets were formulated for prestarter, starter and finisher chickens by following BIS 2007 standards (Table 1).

Table 1: BIS (2007) Standard for broilers

BIS (2007)			
	Pre-starter	Starter	Finisher
CP (%)	23	22	20
ME (kcal/kg)	3000	3100	3200

Table 2: Chemical composition of feed ingredients (% DM basis)

S. No.	Particulars	Maize	Soya-DOC
1	Dry matter	91.07	92.1
2	Crude protein	9	44
3	Crude fibre	2.35	6.3
4	Ether Extract	3.58	1.5
5	Total ash	1.65	2.38
6	Nitrogen free extract	83.42	58.42

The probiotic (encapsulated *Saccharomyces cerevisiae*) was sponsored by Venkateshwara Pvt. Ltd were subjected to chemical analysis in the laboratory at Department of Animal Nutrition, PGIVAS, and Akola.

Dietary Treatment

The experimental birds were subjected to the following dietary treatments.

Table 3: Details of the dietary treatment

Groups	Dietary Treatments No. of Bird in Replicate	No. of Replicate	No. of Birds
T ₀	Standard broiler chicken diet as per BIS, 2007	4	60
T ₁	Standard broiler chicken diet as per BIS, 2007+ oregano essential oil @ 0.15 gm/kg diet	4	60
T ₂	Standard broiler chicken diet as per BIS, 2007+probiotic (encapsulated <i>Saccharomyces cerevisiae</i>) @ 200 gm/ tonnes	4	60
T ₃	Standard broiler chicken diet as per BIS, 2007+oregano essential oil @ 0.15 gm/kg diet+ probiotic (encapsulated <i>Saccharomyces cerevisiae</i>) @ 200 gm/ tonnes T ₁)	4	60
	Total birds	16	240

Total Microbial Count (*E. coli* spp., *Salmonella* spp., *Clostridia* spp.)

Caecal content of the specimens were taken aseptically and were transferred into sterile plastic bags and immediately transported in cold chain to the laboratory. One gram of each sample was diluted 1:9 (wt./vol) in sterile saline. All samples were subjected to 10 sequential dilutions 1:9 (vol/vol), and 0.1 ml of each sample was plated as duplicates by using spread plate method for *E coli*-EMB agar, *Salmonella-shegella* agar and *Clostridium*-nutrient agar. The samples were incubated for 22 ± 2 h at 37°C. Incubation procedure was conducted under aerobic (*E. coli* and *Salmonella*) and anaerobic (*Clostridium*) condition by using incubator. After incubation, typical colonies were counted. The results were expressed as log10 colony-forming units per g of ileal digesta (log10 CFU/g). Results for each bacterium (*E. coli*, *Salmonella* and *Clostridia* spp.) were also checked about its presence.

Statistical Analysis

The results obtained from various parameters *i.e.* blood biochemistry and immune responses were expressed as means ±SE (standard error). The data was analyzed by using Statistical Package for the Social Sciences (SPSS) Version 17.0. The differences between means were subjected to ANOVA by univariate analysis using General Linear Model. A ‘P’ value less than 5% was considered as statistically significant (P<0.05).

Results and Discussion



Total Microbial Count (*E. coli* spp.)

It was observed from the Table 4 that, data pertaining to values of *E. coli* count between the treatment groups were found to be non-significant. The value of *E. coli* count (10^7 CFU/g) for each treatment was found to be 6.34 ± 0.3 , 5.66 ± 0.84 , 5.2 ± 0.79 and 4.35 ± 1 in T₀, T₁, T₂ and T₃ respectively with mean value of 5.38 ± 0.39 . It was observed that Treatment group T₃ had numerically lower value than other groups. Whereas, differences among the treatments T₃, T₂, T₁ and T₀ found to be non-significant. In contrast to the present results Mathlouthi *et al.* (2015) in vitro antimicrobial activities of 3 essential oils- oregano, rosemary and a commercial blend of essential oils (BEO) against pathogenic bacteria *Escherichia coli*, reported significant decrease in the bacterial concentration in the treatment. Total bacterial counts (coliforms particularly) in caecal contents were decreased for birds fed with blend of plant extracts containing oregano, fenugreek, chamomile and fennel decreased Attia *et al.* (2017). Du *et al.* (2015), Sarica *et al.* (2009), Giannenas *et al.* (2016), Manafi *et al.* (2018) also reported similar results.

Total Microbial Count (*Salmonella* spp)

It was observed from the Table 4 that, there was significant differences for *Salmonella* count between the treatment groups. The value of *Salmonella* count for each treatment was found to be 4.6 ± 0.67 , 5.09 ± 0.17 , 4.13 ± 0.62 and 2.76 ± 0.61 in T₀, T₁, T₂ and T₃ respectively with mean value of 4.14 ± 0.31 10^7 CFU/gm. There were significant differences for *Salmonella* count between the treatment groups (Table 4). Treatment group T₃ differ significantly followed by T₂ than T₀ and T₁ while treatment groups T₁ and T₂ differed non-significantly. The lowest value was recorded in T₃ followed by T₂, T₁ and T₀ treatment group respectively. The result of the present study was in accordance with Mathlouthi *et al.* (2015) who fed oregano and rosemary essential oil in broiler and observed and reported decreased salmonella Indiana population in intestine of birds in the treatment groups. Manafi *et al.* (2018) also found the similar results.

Total Microbial Count (*Clostridia* spp)

It was observed from the Table 4 that, the values of *Clostridia* count for each treatment were 2.27 ± 0.02 , 2.05 ± 0.06 , 1.81 ± 0.27 and 1.37 ± 0.34 in T₀, T₁, T₂ and T₃ respectively 10^7 CFU/gm of sample. There were significant differences for *Clostridia* count between the treatment groups (Table 4). Treatment group T₃ differs significantly followed by T₂ than T₀, T₁ while treatment groups T₁ and T₂ differed non-significantly. The lowest value was recorded in T₃ followed by T₂, T₁ and T₀ treatment group respectively.

Table 4: Gut microbes of different dietary treatment

Treatment	<i>E. coli</i> count (10 ⁷ CFU/gm)	<i>Salmonella</i> (10 ⁷ CFU/gm)	<i>Clostridia</i> (10 ⁷ CFU/gm)
T0	6.34 ^a ±0.3	4.6 ^b ±0.67	2.27 ^b ±0.02
T1	5.66 ^a ±0.84	5.09 ^b ±0.17	2.05 ^b ±0.06
T2	5.2 ^a ±0.79	4.13 ^{ab} ±0.62	1.81 ^{ab} ±0.27
T3	4.35 ^a ±1	2.76 ^a ±0.61	1.37 ^a ±0.34
Mean	5.38±0.39	4.14±0.31	1.87±0.12

Treatments in column bearing common superscripts doesn't differ significantly ($P < 0.05$)

The results of the present study are in agreement with Du *et al.* (2015) who showed significantly decrease in the *clostridial* concentration when fed with the active ingredient of oregano oil in the broiler chicken diet.

Conclusion

The results obtained from this study was concluded that the performance of commercial broiler chickens fed with oregano (*Origanum vulgare*) oil as phytobiotic growth promoter with probiotic (encapsulated *Saccharomyces cerevisiae*), has showed a positive impact on GIT and decreased the harmful microbial count of *E. coli*, *Salmonella* and *Clostridia spp* on GIT.

Acknowledgement

The authors are grateful to Director of Research, MAFSU, Nagpur and Associate Dean, Post Graduate Institute of Veterinary and Animal Sciences, Akola for providing necessary facilities.

References

1. Al-Natour MQ and Alshwabkeh KM. 2005. Using varying levels of formic acid to limit growth of *Salmonella gallinarum* in contaminated broiler feed. *Asian-Aust J Anim Sci*, 18 pp. 390-395.
2. AOAC. 2012. Association of official analytical chemist, official method of analysis of AOAC international, 19th Edn. Washington D.C., U.S.A.
3. Attia G, El-Eraky W, Hassanein E, El-Gamal M, Faraha M and Hernandez A. 2017. Effect of dietary inclusion of plant extract blend on broiler growth performance, nutrient digestibility, caecal microflora and intestinal histomorphology. *J. Poult. Sci.* 16 (9): 344-353.
4. Awaad MH, Elmenawey HM and Kawkab AA. 2014. Effect of a specific combination of carvacrol, cinnamaldehyde, and on the growth performance, carcass quality and gut integrity of broiler chickens. *Vet. World*, EISSN: 2231-0916.
5. Barrow PA. 1992. Probiotics for chicken (In Probiotics: The scientific basis, ed. by R. Fuller). Chapman and Hall, London. pp 225-257.
6. BIS 2007. Nutrient Requirements of Poultry. Bureau of Indian Standards, 5th revision. New Delhi, India.
7. Cenesz S, Yaman H, Ozcan A and Karademr G. 2008. Effect of kefir as probiotic on serum cholesterol, total lipid, aspartate amino transferase and alanine amino transferase activities in broiler chicks. 72(2): 61-67.
8. Chitra P, Mohan B and Vishwanathan K. 2004. Effect of probiotic with ascorbic acid on growth performance of broilers in the summer season. *Ind. J. Poult. Sci.* 39(3): 281-284.

9. Du E. 2016. Effects of thymol and carvacrol supplementation on intestinal integrity and immune responses of broiler chickens challenged with *Clostridium perfringens*. *Journal of Anim. Sci. Biotechnol.* 7:19–29.
10. Edwards JR. 2002. Studies on the efficiency of cholecalciferol and derivatives for Stimulating phytate utilization in broilers. *Poult. Sci.*, 81(7):1026-1031.
11. FAO/WHO. 2002. Guidelines for the evaluation of probiotics in food, Food and Agriculture Organization of the United Nations and World Health Organization Working Group Report, London Ontario, Canada).
12. Gao J, Zhang HJ, Yu SH, Wu SG, Yoon I, Quigley J, Gao YP and Qi GH. 2008 Effects of yeast culture in broiler diets on performance and immune modulatory functions, *Poult. Sci.* 87: 1377-1384.
13. Giannenas I, Athina T, Ioannis S, Achilleas K, Stylianos S, Nikolaos P, Ioannis A, Ioannis S. 2016. The effectiveness of the use of oregano and laurel essential oils in chicken feeding. *Ann. Anim. Sci.*, 16(3):779–796.
14. Gopi M, Kumaragurubaran K, Haranahalli VM, Paramasivam T, Manickam K, Moorthy D, Bharemara LB, Manika RP. 2013. Essential oils as a feed additive in poultry nutrition. *Advan. in Anim. and vet. Sci.* 2(1):40.
15. Haldar S, Ghosh T, Toshiwati K and Bedford MR. 2011. Effects of yeast (*Saccharomyces cerevisiae*) and yeast protein concentrate on production performance of broiler chickens exposed to heat stress and challenged with *Salmonella enteridis*. *Anim. Feed Sci. Technol.* 16:61–71.
16. Line EJ, Bailey SJ, Cox NA, Stern NJ and Tompkins T. 1998. Effect of yeast-supplemented feed on *salmonella* and *campylobacter* populations in broilers. *Poult. Sci.* 77:405-10.
17. Luna G. 1968. Manual of histological staining methods of the armed forces Institute of Pathology, 3rd ed. New York: McGraw-Hill Book Co., 258 pp.
18. Manafi M, Hedayati M and Khalaji S. 2016. Effectiveness of phytogenic feed additive as alternative to bacitracin methylene disalicylate on hematological parameters, intestinal histomorphology and microbial population and production performance of Japanese quails. *Asian-Australas. J. Anim. Sci.* 29(9): 1300-1308.
19. Mead GC. 2000. Prospects for “competitive exclusion” treatment to control salmonellas and other foodborne pathogens in poult. *Vet. J.* 159:111–123.
20. Mathlouthi N, Bouzaienne T, Oueslati I, Recoquilly F, Hamdi M, Urdaci M and Bergaoui R. 2012. Use of rosemary, oregano, and a commercial blend of essential oils in broiler chickens: In vitro antimicrobial activities and effects on growth performance, *J. Anim. Sci.* 90:813–823.
21. Chaudhary N, Saikia B, Dowarah R, Tamuly S and Sapkota D. 2017. Effect of feeding curd as a probiotic on growth performance, nutrient utilization, blood biochemical and ceecal microbial profile in broilers. *International Journal of Livestock Research*, 7(12): 165-173. <http://dx.doi.org/10.5455/ijlr.20170329100757>.
22. OIE. 1992. Manual of standards for diagnostic test and vaccines off Int. Epizootics, Paris.
23. Owings WJ, Reynolds DL, Hasiak RJ and Ferket PR. 1989. Influence of dietary supplements with *Streptococcus faecium* M-74 on broiler body weight, feed conversion, carcass characteristics, and intestinal microbial colonization. *Poult. Sci.* 69:1257–1264.
24. Paryad A and Mahmoudi M. 2008. Effect of different levels of supplemental yeast (*Saccharomyces cerevisiae*) on performance, blood constituents and carcass characteristics of broiler chicks. *Afr. J. Agric., Res.* 3(12): 835-842.
25. Pelicano ERL, Souza PA, Souza HBA, Leonel FR, Zeola NMBL, Boiogo MM. 2005. Productive traits of broiler chickens fed diets containing different growth promoters. *Revista Brasileira de Ciência Avícola.* 6 (3):177-182.
26. Pietras M. 2001. The effect of probiotic on selected blood and meat parameters of broiler chickens. *J. Anim. Feed. Sci.* 10 (suppl 2):297-302.
27. Saad S, Abbod M and AboYones A. 2014. Effects of some growth promoters on blood hematology and serum composition of broiler chickens. *Int. J. Agri. Research*, 9(5): 265-270.

28. Shanmuga PB. and Saravana BS. 2013. Effect of different levels of supplemental probiotics (*saccharomyces cerevisiae*) on performance, haematology, biochemistry, microbiology, histopathology, storage stability and carcass yield of broiler chicken. Department of Botany, C. N. College, Erode-638004, Tamil Nadu, India.
29. Sarica S, Corduk M, Yarim GF, Yenisehirli G and Karatas U. 2009. Effects of novel feed additives in wheat-based diets on performance, carcass and intestinal tract characteristics of quail. *South Afr. Journal of Anim. Sci.* 39(2).
30. Tiihonen K, Kettunen H, Bento MHL, Saarinen M, Lahtinen S, Ouwehand AC, Schulze H and Rautonen N. 2010. The effect of essential oils on broiler performance and gut microbiota. *Bri. Poult. Sci.* 51:381–392.
31. Yalçın S, Eser H, Yalçın S, Cengiz S and Eltan. 2013. Effects of dietary yeast autolysate (*Saccharomyces cerevisiae*) on performance, carcass and gut characteristics, blood profile, and antibody production to sheep red blood cells in broilers. *J. Appl. Poult. Res.* 22 :55–61.