

# Epidemiology, Risk Factors and Economics of Echinococcosis in India: A Review

**Kushal Grakh, Anand Prakash, Dinesh Mittal\*, Pankaj Kumar and Ramesh Kumar**

Department of Veterinary Public Health and Epidemiology, Lala Lajpat Rai University of Veterinary and Animal Sciences, Hisar, Haryana, INDIA

\*Corresponding Author: [mittalvet@luvas.edu.in](mailto:mittalvet@luvas.edu.in)

**How to cite this paper:** Grakh, K., Prakash, A., Mittal, D., Kumar, P., & Kumar, R. (2020). **Epidemiology, Risk Factors and Economics of Echinococcosis in India: A Review.** *International Journal of Livestock Research*, 10(7), 1-10. doi: <http://dx.doi.org/10.5455/ijlr.20200418070900>

**Received** : Apr 18, 2020  
**Accepted** : Jun 06, 2020  
**Published** : Jul 31, 2020

Copyright © Grakh *et al.*, 2020

This work is licensed under the Creative Commons Attribution International License (CC BY 4.0). <http://creativecommons.org/licenses/by/4.0/>



**Open Access**

## Abstract

*Echinococcosis caused by cestode of genus Echinococcus is an important zoonotic disease with worldwide distribution. Two of the important echinococcosis are cystic echinococcosis (CE) and alveolar echinococcosis (AE) caused by metacestodes of Echinococcus granulosus and E. multilocularis, respectively. The definitive hosts for CE or hydatidosis are carnivores and intermediate hosts are herbivores, with humans acquiring the accidental infection from carnivores. The larval stages of E. granulosus can parasitize internal organs of animals and humans and as a result responsible for substantial morbidity and mortality. Worldwide, India is ranked first in CE cases amounting to 12% of global CE cases. Apart from morbidity and mortality in humans and animals, the economic losses in animals arise from production losses, condemnation of cyst affected carcasses, and losses from poor reproductive performances. The data regarding the losses in animals is often underreported. In humans, 8,71,000 disability- adjusted-life- years (DALY) globally each year points toward the severity of the disease. The various risk factors such as illegal slaughter/ slaughter in uncertified slaughterhouses, large population of stray dogs, dependency of rural population to raise livestock and practice of feeding dogs with offal of home butchered animals facilitate CE transmission in India, which makes it a public health, veterinary and social challenge. Systematic and national level surveillance of disease in rural livestock and human population, awareness programmes about its zoonotic nature, coordinated efforts from veterinary and medical science for epidemiology and diagnosis, and strict legislation in post mortem inspection at abattoirs is needed for prevention and control of the disease.*

**Keywords:** Abattoir, Echinococcosis, Host, Zoonosis

## Introduction

Echinococcosis is a neglected zoonotic parasitic infection caused by the larval stage of tapeworm parasite belonging to the genus *Echinococcus*. The life cycle of this tapeworm is sustained between dogs (definitive host) and herbivores (intermediate host). Humans can also develop the disease owing to consumption of food stuff contaminated with dog faeces. *Echinococcus granulosus* causes cystic echinococcosis (CE) whereas *Echinococcus multilocularis* is responsible for alveolar echinococcosis (AE). AE is non-endemic in India and information about its clinical cases, prevalence and epidemiology is limited (Madhusudhan *et al.*, 2016). However, CE is an endemic zoonosis in many parts of the world including India. The larval stages of *E. granulosus* can parasitize liver, lung, lung and other organs of humans, cattle, buffalo, sheep, pigs and other animals (Haridy *et al.*, 2006). It is recognized as one of the seventeen neglected human diseases in the world with more than one million human cases (WHO, 2018). India is ranked first in CE cases with 12% (119,320 cases) reported from India out of 973662 global human CE cases (Hotez and Damania, 2018). Because of the involvement of canines, herbivores and human, the CE is the disease of high public health, veterinary and socioeconomic importance in India as well as the world.

Echinococcosis persists mainly in domestic cycle involving domestic dog as definitive host and various species of domestic animals as intermediate hosts. The *E. granulosus* eggs can survive under warm, humid and cold climatic conditions for weeks to months, but sensitive to desiccation (Eckert *et al.*, 2003). CE is distributed worldwide in human and animal population with highest prevalence found in parts of Eurasia, Africa, Antarctica and South America (Eckert and Deplazes, 2004). However, Iceland and Greenland islands are free from *E. granulosus* and only sporadic cases were detected from others such as New Zealand, Tasmania and Southern Cyprus (Eckert and Deplazes, 2004). Echinococcosis or hydatid cysts have been found in cattle, buffalo, sheep and other food producing animals in India. A total annual loss of Rs. 11.47 billion in India is estimated with cattle and buffalo population affected with CE and this itself is an understatement owing to under reporting of the disease in animals (Singh *et al.*, 2014a). However, the losses due to surgical interventions, loss of productivity and hospital due to CE in humans were estimated to be Rs. 472.72 million (Singh *et al.*, 2014a). Unplanned or illegal slaughtering, large population of stray dogs, improper disposal of dead animals and close association of rural population with animals increase the possibility of disease transmission to humans. Latest estimates on CE suggests annual global incidence of at least 1,88,000 new CE cases (Rojas *et al.*, 2018). Around 12% of the global CE cases are from India which is because of favorable climatic conditions such as warm and humid climate, availability of large numbers of intermediate host dependent on grazing activities, lack of awareness and poor collaboration between veterinary and medical sciences in India. The present article emphasizes the epidemiological aspects, risk factors and economic losses associated with the disease.

## Life Cycle

The life cycle of these parasites is indirect and requires two mammalian hosts i.e. definitive and intermediate hosts. The life cycle starts with the adult stage of the parasite in small intestine of the dogs or other canids (definitive host). The definitive host pass the parasitic eggs in faeces which contaminate the soil, pasture, vegetable gardens and water sources like ponds in the environment. In the environment the eggs can remain viable for weeks to months up to a wide temperature range of 38°C to -30°C, however infectivity tends to decrease as the temperature increases (NCDC, 2016). After ingestion of contaminated food stuff by intermediate hosts (domestic or wild ungulates), eggs hatch in small intestine and release larval stage (oncosphere) that penetrates the intestinal wall. Through intestinal blood vessels, the oncosphere reaches various organs, especially liver and lungs. In these organs, the oncosphere develops into second stage known as metacestode which are cystic structure, typically filled with clear fluid (hydatid fluid) that enlarge to produce protoscolices and daughter cysts. These cysts infesting various visceral organs of the intermediate host are ingested by the definitive host. In the small intestine of definitive host protoscolices evaginate, attach to intestinal mucosa and develop into adult stages in 32 to 80 days thereby completing the life cycle (Thompson and Lymbery, 1990). Humans acquire the infection through accidental ingestion of the eggs, which in turn develop into the parasite's larval stage (metacestode) in internal organs and ultimately CE. The exact time required for development of various stages of cyst in humans is not known, but it is estimated to be more than 10 months post infection (Eckert and Deplazes, 2004). Human act as dead-end host in the disease transmission cycle.

## Clinical Signs

The disease is primarily asymptomatic in definitive and intermediate animal hosts. In humans, the disease may

remain asymptomatic for years, and signs and symptoms may appear subject to the organ affected and severity of infection (WHO, 2018). However, irrespective of the location of cyst, the primary symptoms in affected human population include pain in abdomen, followed by lump in abdomen (Rao *et al.*, 2012). In addition, the symptoms may vary depending upon the localization of the cyst (Sharma *et al.*, 2013).

## **Predisposing Factors**

### **Sex/Gender**

The conflicting reports exist indicating sex as a predisposing factor for the disease in humans and animals. Higher rate of infection in females have been observed in goats and it is possible because of longer survival period of parasite in females due to potential link between sexual hormones and response of the immune system (Blancas *et al.*, 2007; Godara *et al.*, 2014). Other possible reason might be that female animals are slaughtered at older age as they are retained for reproductive purposes and therefore longer life expectancy provides more chances of exposure to the infection (Ibrahim, 2010). Similarly, in humans higher cases are observed in females as compared to males (Rao *et al.*, 2012). The reasons might be due to the fact that in rural communities, females are more involved in livestock management activities and hence have more chance to acquire the disease from environmental contamination (Singh *et al.*, 2014a). On contrary, some hospital-based seroprevalence studies in humans and abattoir-based studies in animals have not found any significant differences in gender predisposition (Khurana *et al.*, 2007; Zaman *et al.*, 2017; Ganaie *et al.*, 2018). So, no consensus can be found or established regarding predisposition of sex in the disease.

### **Age Group**

Incidence of CE is higher in older animals as compared to younger ones and this finding is consistent with animal population viz. cattle, sheep, goat, pig, buffaloes (Singh *et al.*, 2012; Otero-Abad and Torgerson, 2013). A study carried out in sheep and goats recorded 1.6 times higher hydatidosis prevalence in older sheep and goat (more than three years old) as compared to young ones (Marshet *et al.*, 2011). It could be because most female sheep and goats, which are retained for reproductive purposes, are sent for slaughter at old age and therefore diagnosed at older age as compared to young ones. In humans, most commonly affected age group ranges from 17-50 years (Rao *et al.*, 2012; Gochhait *et al.*, 2015). It may be due to the fact that as infection remain asymptomatic in animals and humans and also parasite has a long incubation period, hence, most of the cases are diagnosed at an older age (Craig *et al.*, 2017).

### **Seasonal Pattern**

Seasonal influence has been the least studied parameter and does not seem to have any significant effect on the disease prevalence among intermediate hosts (Ganaie *et al.*, 2018).

### **Organ Predilection**

Irrespective of the species of intermediate animal host, liver and lungs are observed as most commonly affected or predilected area for hydatid cyst formation. Liver as well as lungs are considered as most affected organs in animals (Pednekar *et al.* 2009; Singh *et al.*, 2012; Godara *et al.*, 2014; Sajjan *et al.*, 2015). Other organs involved are spleen, heart, kidney, mesentery and diaphragm. In humans, liver is found to be most affected organ followed by lungs. Rare cases of skeletal cysts are also found in humans (Rao *et al.*, 2012; Sharma *et al.*, 2013).

### **Risk Factors for the Disease**

The suitable conditions for the establishment and transmission of Echinococcosis exist in India such as large population of intermediate hosts, close contact of human-dog and dog and intermediate host, browsing and grazing on contaminated pasture, warm and humid and cold climate conditions during different part of seasons (Singh *et al.*, 2014a). India has a huge livestock population of 536.76 million, out of which 536.16 million can act as intermediate hosts to complete the life cycle (20<sup>th</sup> Livestock Census, 2019). These intermediate hosts graze on pastures contaminated with dog faeces and ingest the echinococcal eggs. High number of stray dog population in both urban and rural areas in India is another risk factor responsible for higher prevalence of *E. granulosus* (Table

1). These stray dogs receive no or an inadequate treatment with anti-parasitic medications and thus they play an important role in the transmission of the disease in the country. Open dumping of dead animals in rural part of the country with an easy access to stray dogs further complicate the problem. Pet dogs as guarding animals, dog owners, animal handlers, veterinarians and laboratory workers are also at higher risk of infection, since the eggs can contaminate water, fruits and vegetables. These eggs can stick to the fur of an animal and get transferred from hands to the mouth of humans in absence of personnel hygiene. Shephard dogs have been found to be at higher infection risk as compared to stray dogs, due to their free access to offals and other animal wastes (Otero-Abad and Torgerson, 2013). Frequent contact of rural human population with animals due to their dependency on livestock for income, transportation, cultivation etc. is another risk factor. Other factors like lack of awareness about the parasite, inappropriate meat inspection at slaughterhouses and offal disposal by illegally run abattoirs also contributes to domestic cycle of transmission.

**Table 1:** Prevalence (%) of *E. granulosus* in stray dog population in India

Location/State	Prevalence	Reference
Kurnool, Andhra Pradesh	33.3	Reddy <i>et al.</i> , 1958
Delhi and Uttar Pradesh	14	Singh and Dhar, 1988
Assam	17.02	Deka <i>et al.</i> , 2008
Mizoram	18.18	Deka <i>et al.</i> , 2008
Meghalaya	27.77	Deka <i>et al.</i> , 2008
Bangalore urban, Karnataka	4.35	Prathiush <i>et al.</i> , 2008
Maharashtra	5.22 - 6.57	Nikale <i>et al.</i> , 2014; Ingole <i>et al.</i> , 2018
Gurugram, Haryana	13.5	Varma, 1990

### Prevalence of *E. granulosus*

Accurate diagnosis and prevalence of *Echinococcus* infection in the intermediate and definitive hosts is an important component for establishing echinococcosis and preventing human and livestock infection (Moon and Khemalpure, 2017). The already published data related to echinococcosis in dog (Table 1) and livestock population (Table 2) indicates significant prevalence of disease in parts of India. As per 20<sup>th</sup> livestock census, 2019, the total dog population is 9.43 million in India. But according to other animal welfare groups and websites the country is home to about 30 million stray dogs, which amounts to 1 stray dog per 42 people in the country (Socialcops, 2020; Statista, 2020). From the various published reports, (Table 1) the range of *E. granulosus* infection in stray dog population vary from 4.35% in Karnataka to 33.3% in Andhra Pradesh in India.

The available data indicate that cattle, buffalo, sheep, goat and pig play an important role as intermediate hosts in India. The higher prevalence in all the intermediate hosts might be due to the grazing of these animals on contaminated pastures or by drinking dog feces contaminated water (Otero-Abad and Torgerson, 2013). Since CE is asymptomatic in animals, abattoir-based studies relied for prevalence data in domestic livestock animals. Most of these studies took place in large abattoirs where animals are brought from random places that are intensively managed or large-scale production system. The studies involving urban based abattoirs, hence may indicate irrelevant diseases prevalence in rural communities, which is expected to be significantly higher (Pednaker *et al.*, 2009). Most of studies reportedly claiming prevalence of CE in specific region of India (north, south, east and west) are based upon the abattoir data in that particular location or region, hence such studies may not actually reflect true prevalence and actual prevalence is expected to be higher.

The seroprevalence against CE have shown increasing trends in India. An overall significant increase from 15.0% (1984-2003) to 28.6% seropositivity (2004-2015) in humans has been observed in India (Khurana *et al.*, 2007; Zaman *et al.*, 2017). WHO has recommended the use of clinical and laboratory data of hospital cases as practicable indicator of the incidence of the disease in humans.

**Table 2:** Zone and state wise prevalence (%) of CE in intermediate hosts

Region	Prevalence					Literature
	Cattle	Buffalo	Sheep	Goat	Pig	
<b>Northern India</b>						
Haryana			4.7	1.9	1.3	Varma, 1990
Punjab	7.6	8.9-50.96	5.1	2.8	4.66	Deka <i>et al.</i> , 1983, Sharma <i>et al</i> 2004, Jadhav <i>et al.</i> , 2013, Khan <i>et al.</i> , 2013
HP			7.2	2.3		Jitendran, 1996
J&K				19.8		Godara <i>et al.</i> , 2014
Uttar Pradesh		14.82- 36	2.56-6	1.45-2	0.9-1.42	Deka and Gaur 1998, Varma and Malviya 1992, Irshadullah <i>et al.</i> , 1989; Ganaie <i>et al.</i> , 2018
Delhi			18.5			Rana <i>et al.</i> , 1986
Diff. Location	5.39	4.36	2.23	0.41	3.09	Singh <i>et al.</i> , 2014b
Diff. Location		48.1	30.50%	21		Singh and Dhar, 1988
Range	5.4-7.6	4-51	2.2-30	0.41-21	0.9-4.6	
<b>Western India</b>						
Maharastra	5.1	3.81	0.02-1.06	0.3	0.87	Pednekar <i>et al.</i> , 2009, Gatne 2001
Rajashthan			6.57	4.12		Shekhawat <i>et al.</i> , 2005
MP		1.19				Jatav and Garg, 2012
Range	5.1	1.2-3.8	0.02-6.6	0.3-4.1	0.9	
<b>Southern India</b>						
AP			5.18-7.05	1.39		Pillai <i>et al.</i> , 1986; Hafeez <i>et al.</i> , 1994
Chennai			5.6-22.4	7-25.8		Sangaran <i>et al.</i> , 2014; Shanmugam <i>et al.</i> , 1994
Karnataka			0.37-4.9	4.78		Vijayasmitha <i>et al.</i> , 1993
Kerala	35.47	33	2.55	1.05		Abraham <i>et al.</i> , 1980
Diff. Location	7.1	9.4	7		11.5	Hafeez, 1997
Range	7.1-35.4	9.4-33	0.4-22.4	1.0-25.8	11.5	
<b>Eastern India</b>						
Assam	16	6.52		2.24-4.9	0.43-1.8	Sarma <i>et al.</i> , 2000; Deka <i>et al.</i> , 2008
Bihar				8.48	7.6-8.25	Kumar <i>et al.</i> , 2007; Prasad, 1981
Meghalaya	21.40%				0.34	Deka <i>et al.</i> , 2008
Puduchery			47.6	37.8		Das and Sreekrishnan, 1998
Sikkim			50	33.3		Katiyar and Sinha, 1981
West Bengal			10.5	3.9	8	Biswas <i>et al.</i> , 1989; Das and Das, 1998
Range	16-21.4	6.5	10.5-50.0	2.2-57.8	0.3-8.3	
Overall	1.6-35.4	1.2-51	0.02-50	1.05-57.8	0.3-11.5	

### Molecular epidemiology of *Echinococcus* species

Mitochondrial DNA sequences have identified 10 different genetic types of *E. granulosus* from G1 to G10 (Craig *et al.*, 2007). Livestock from eastern India demonstrated the presence of four genotypes G1 (sheep strain), G2 (Tasmanian sheep strain), G3 (buffalo strain) and G5 (cattle strain) (Bhattacharya *et al.*, 2007). In northern India, 77.7% of isolates from cattle, pig, buffalo and goat were grouped under G3 strain and 22.2% isolates from sheep are grouped under G1 strain (Singh *et al.*, 2012). Four different genotypes i.e. G1, G2, G3 and G5 are demonstrated in food producing animals viz. cattle, buffalo, sheep and pigs in western India (Pednekar *et al.*, 2009). G5 (*E.*

*ortleppi*) is only reported in India (Pednekar *et al.*, 2009). The abattoir-based studies in animals indicate that G3 is most common strain of *E. granulosus* circulating in livestock in India followed by G5, G1 and G2 strain.

The strains of *Echinococcus* spp. circulating in livestock in India are well adapted to cause infection in all the intermediate hosts of *E. granulosus* including humans, thus proving to be of zoonotic importance. Globally, most of the human cases of CE are found to be infected with G1 strain of *E. granulosus* (Moro and Schantz, 2009). Despite the isolation of G5 and G6 in humans, G1 and G3 remained commonly encountered strains in human CE in India (Sharma *et al.*, 2013). The wide variation in *E. granulosus* strains may influence the life-cycle patterns, development rate, specificity to host, transmission, antigenic proteins in fluid and response to treatment (Craig *et al.*, 2007).

### Diagnosis

In the intermediate hosts, diagnosis of the disease depends on the meat inspection or post-mortem detection of the larval cyst form which can occur in almost any organ particularly in the liver and lungs (Siles-Lucas *et al.*, 2017). In live animals, ultrasonography and radiography have been used previously to detect the presence of cysts in lungs and abdominal cavity. However, demonstration of scolices, hooklets or protoscolices in aspirated fluid (fine needle aspiration/ultrasound guided aspiration) by direct microscopic examination or stained smears is definitive diagnosis of cystic echinococcosis (Singh *et al.*, 2014a). Molecular methods especially PCR are used widely to characterize *Echinococcus* spp. nowadays and proved to be of great value in identification of *Echinococcus* spp. in samples collected from livestock at abattoirs for epidemiological purposes (Cardona and Carmena, 2013). Serological tests using ELISA are not much reliable in diagnosis of echinococcosis in intermediate hosts (Siles-Lucas *et al.*, 2017). The diagnosis of echinococcosis in definitive hosts (dogs or other carnivores) requires the demonstration of the adult cestodes of *Echinococcus* spp. in their faeces or small intestine or detection of specific coproantigens (using ELISA) or copro DNA (using specific primer-based PCR amplification). Sedimentation and counting technique (SCT) for eggs in feces of canines are regarded as the 'gold standard' for assessing the sensitivity and specificity of other techniques (Eckert *et al.*, 2003). However, the copro-DNA (PCR) test has a greater sensitivity than SCT.

In humans, non-invasive imaging techniques such as CT scans, MRI and ultrasonography (USG) are used for detecting and defining the extent and condition of avascular fluid-filled cysts in most organs. CT and USG are widely used for screening, clinical diagnosis, and monitoring of treatment of lungs and liver and intra-abdominal cysts respectively (Giri and Parija, 2012). Radiography can also be used in the cases where cyst calcification is suspected. MRI is very useful tool for diagnosis of rare cerebral hydatid cysts and helpful in diagnosing multiple cysts and cyst capsules (Bukte *et al.*, 2004). However, serological tests (ELISA and indirect hemagglutination test), are highly sensitive, but specific confirmation can be obtained by detecting echinococcal antigens by immunodiffusion (arc 5) or immunoblot assays (8-, 21 –kD bands) (Eckert *et al.*, 2003). Counter current immunoelectrophoresis (CIEP) and Casoni's test have been previously used as diagnostic test for hydatid cyst, but not common nowadays (Giri and Parija, 2012). The urine samples are also used for antigen detection as an alternative or in addition to serum because of the non-invasive nature of sample collection and comparable sensitivity and specificity (Sunita *et al.*, 2011). Co-agglutination tests has also been found cheap and rapid using urine and serum samples for detection of hydatid antigen (Parija and Shivaprakash, 2000).

### Economics of Echinococcosis

Total livestock population of India is 536.76 million (20<sup>th</sup> Livestock Census-2019). All species except yak, mule, donkey, horse and ponies in India act as intermediate host for echinococcosis. Diseases or infections act as negative input to the production of animal which affect the efficiency of livestock and thus adds to losses (Thrusfield, 2007). The economic losses associated with hydatid disease can be in the form of poor growth, loss of benefits due to condemnation of cyst affected offal mainly liver, production losses by decrease in weight, milk production, wool and fleece/hide value, and reproductive performance (Vaidya *et al.*, 2014). Cost incurred in diagnosis, hospitalization, treatments including surgery, income loss, temporary or permanent disability or impairment and post-operative fatalities in humans which reduces the quality life, throw the light on public health and economic significance of CE. Echinococcosis is estimated to be the cause of 19,300 deaths and around 8,71,000 disability-adjusted life-years (DALYs) globally each year (WHO, 2018). The attempts in food producing animals have been made to calculate the economic cost due to organ condemnation. In Maharashtra, echinococcosis lead to economic losses to the tune of Rs 26, 78, 721 per year (Vaidya *et al.* 2014) due to offal and carcass condemnation. The

economic loss due to the condemnation of edible offal in pigs can be as high as Rs. 11,396.10 in some states (Sajjan *et al.* 2015). The systematic analysis of economic losses based on available literature in livestock and humans in India estimated median economic losses to the amount of Rs. 11.47 billion (approx. US \$ 212.35 million) where cattle and buffalo industry shared most of the losses i.e. 93.05% of total livestock and 88.88% of the livestock and human losses. Human echinococcosis associated losses are estimated to be Rs. 472.72 million which are likely to be under-estimated due to under-reporting of the disease in the country (Singh *et al.*, 2014a).

## Conclusion

It can be concluded that CE has significant economic and public health impact worldwide. India have highest number of CE cases (12% of global cases) worldwide. This figure is likely to go up as the disease is often underreported in India. However, the advent of technology, developing health infrastructure, and improved research facilities in the country have facilitated CE diagnosis. In order to aid disease control and prevention, the active surveillance of the disease in livestock and human is needed. The extension programmes about basic hygiene and zoonotic nature of the disease, legalization of slaughter house and proper meat inspection, and combined efforts from medical and veterinary sciences are must to control and prevent disease- associated morbidity and mortality

## Conflict of Interests

There is no conflict of interest.

## Publisher Disclaimer

IJLR remains neutral concerning jurisdictional claims in published institutional affiliation.

## References

- 20<sup>th</sup> Livestock Census, 2019. Basic Animal Husbandry Statistics. Department of Animal Husbandry, Dairying and Fisheries, Government of India.
- Abraham, J., Madhvan P.K. and Iyer, R.P. (1980). Incidence of hydatidosis in animals slaughtered in Kerala. *Kerala Journal of Veterinary Sciences*, 11: 247-251.
- Bhattacharya, D., Bera, A.K., Bera, B.C., Maity, A. and Das, S.K. (2007). Genotypic characterization of Indian cattle, buffalo and sheep isolates of *Echinococcus granulosus*. *Veterinary Parasitology*, 143: 371–374.
- Biswas, G., Sen, G.P., Thapa, D. and Lahkar, A. (1989). Hydatidosis in animals in Calcutta. *Indian Veterinary Journal*, 66, 78–80.
- Blancas, M.M., Herrera, E.R., Rodriguez, P.C., Tavizon, J.P., Mercado, R.M. (2007). Gender as a factor of susceptibility to infection in experimental hydatidosis. *Revista Latinoamericana Microbiologia*, 49, 31–37.
- Bukte, Y., Kemalolu, S., Nazaroğlu, H., Ozkan, U., Ceviz, A., Simsek, M. (2004). Cerebral hydatid disease: CT and MR imaging findings. *Swiss Medical Weekly*, 134, 459-67.
- Cardona, G. A., and Carmena, D. (2013). A review of the global prevalence, molecular epidemiology and economics of cystic echinococcosis in production animals. *Veterinary Parasitology*, 192, 10-32.
- Craig, P.S., Hegglin, D., Lightowers, M.W., Torgerson, P.R. and Wang, Q. (2017). Echinococcosis: control and prevention. In *Advances in parasitology* (Vol. 96, pp. 55-158). Academic Press.
- Craig, P.S., McManus, D.P., Lightowers, M.W., Chabalgoity, J.A. and Garcia, H.H. (2007). Prevention and control of cystic echinococcosis. *Lancet Infectious Disease*, 7, 385– 394.
- Das, S.S. and Sreekrishnan, R. (1998). Prevalence of hydatidosis in sheep and goats in Pondicherry. *Journal of Veterinary Parasitology*, 12, 145-146.
- Das, U. and Das, A.K. (1998). Cystic hydatidosis of food animals in greater Calcutta. *The Indian Veterinary Journal*, 75, 387-388.
- Deka, D.K. and Gaur, S.N.S. (1998). – Some studies on the occurrence of hydatidosis in western Uttar Pradesh. *Journal of Veterinary Parasitology*, 12, 43-45.

13. Deka, D.K., Saidul, I., Manoranjan, B., Abdus, S., Isfaqul, H., and Krishna, N. (2008). Prevalence of *Echinococcus granulosus* in dogs and hydatidosis in herbivores of certain north eastern states of India. *Journal of Veterinary Parasitology*, 22, 22-26.
14. Deka, D.K., Shrivastava, G.C., Chhabra, R.C. (1983). Incidence of hydatidosis in ruminants. *Indian Journal of Animal Science*, 53, 200–202.
15. Eckert, J. and Deplazes, P. (2004). Biological, epidemiological, and clinical aspects of echinococcosis, a zoonosis of increasing concern. *Clinical Microbiology Reviews*, 17, 107-135.
16. Eckert, J., Gemmell, M.A. and Soulsby, E.J. (2003). FAO/UNEP/WHO Guidelines for Surveillance, Prevention and Control of Echinococcosis/Cystic Echinococcosis. Geneva: World Health Organization.
17. Ganaie, Z., Samanta, S. and Battoo, A. (2018). Prevalence and associated epidemiological factors of hydatidosis in buffaloes in and around Bareilly, Uttar Pradesh, India. *International Journal of Livestock Research*, 8, 327-334.
18. Gatne, M. L. (2001). Studies on the antigenic profiles of hydatids in farm animals with particular emphasis on specific immunodiagnosis. Ph.D. thesis approved by Dr. Balasaheb Sawant Konkan Krishi Vidyapeeth, Dapoli (Maharashtra).
19. Giri, S., and Parija, S.C. (2012). A review on diagnostic and preventive aspects of cystic echinococcosis and human cysticercosis. *Tropical Parasitology*, 2, 99-108.
20. Gochhait, D., Dey, P., Rajwanshi, A., Nijhawan, R., Radhika, S. and Gupta, N. (2015). Spectrum of fungal and parasitic infections on fine needle aspiration cytology. *Diagnostic Cytopathology*, 43, 450-455.
21. Godara, R., Katoch, R. and Yadav, A. (2014). Hydatidosis in goats in Jammu, India. *Journal of Parasitic Diseases*, 38, 73–76.
22. Hafeez M. (1997). Epidemiology and transmission of cystic echinococcosis: India. *Archive of Parasitology*, 32, 54-55.
23. Hafeez, M. D., Reddy, P. R., Hasina, S., Prasad, K. L. G., Nirmala Devi, K. and Thayered, M. D. (1994). Fertility rate of hydatidosis in cattle, buffaloes, sheep and pigs. *Indian Journal of Animal Science*, 64, 46-47.
24. Haridy, F.M, Ibrahim, B.B., Elshazly, A.M., Awad, S.E., Sultan, D.M., El-Sherbini, G.T., Morsy, T.A. (2006). Hydatidosis granulosus in Egyptian slaughtered animals in the years 2000-2005. *Journal of Egypt Society of Parasitology*, 36, 1087-1100.
25. Hotez, P.J. and Damania, A. (2018). India's neglected tropical diseases. *PLoS Neglected Tropical Disease*, 12, e0006038.
26. Ibrahim, M. (2010). Study of cystic echinococcosis in slaughtered animals in Al Baha region, Saudi Arabia: Interaction between some biotic and abiotic factors. *Acta Tropica*, 113, 26–33.
27. Ingole, R.S., Khaksea, H.D., Jadhava, M.G., Ingole, S.R. (2018). Prevalence of Echinococcus infection in dogs in Akola district of Maharashtra (India) by Copro- PCR. *Veterinary Parasitology: Regional Studies and Reports*, 13, 60–63.
28. Irshadullah, M., Nizami, W.A., Macpherson, C.N. (1989). Observations on the suitability and importance of the domestic intermediate hosts of Echinococcus granulosus in Uttar Pradesh, India. *Journal of Helminthology*, 63, 39–45.
29. Jadhav, S.K., Sathapathy, S., Gohain, S. and Joshi, S.K. (2013). Prevalence of hydatidosis in buffaloes. *Research Journal of Animal Veterinary and Fishery Sciences*, 1, 23-25.
30. Jatav, G. P. and Garg, U. K. (2012). Hydatidosis in the livers of buffaloes (*Bubalus bubalis*) in the Malwa region of Madhya Pradesh. *Buffalo Bulletin*, 31, 185-188.
31. Jithendran, K. P. (1996). Occurrence of hydatidosis and various liver fluke infestation in sheep and goat in Kangra Valley: An abattoir Study. *Journal of Veterinary Parasitology*, 10, 63-67.
32. Katiyar, R. D. and Sinha, A. K. (1981). Hydatid disease in livestock in Sikkim. *Livestock Advisor*, 6, 57-58.
33. Khan, A.M., Gazi, M. and Bashir, S. (2013). Seasonal prevalence of hydatidosis in buffaloes –A retrospective study. *Veterinary World*, 6, 647-650.
34. Khurana, S., Das, A. and Malla, N. (2007). Increasing trends in seroprevalence of human hydatidosis in North India: a hospital-based study. *Tropical Doctor*, 37, 100-102.

35. Kumar, S., Quasim, P., Kaushik, P. and Samantaray, S. (2007). Prevalence of *Echinococcus granulosus* cysts in food animals in and around Patna. *Journal of Veterinary Public Health*, 5, 25-28.
36. Madhusudhan, K.S., Srivastava, D.N., Dash, N.R., Venuthruimilli, A., Sharma, R., Gamanagatti, S. and Gupta, A.K. (2016). Alveolar echinococcosis of the liver: a diagnostic problem in a nonendemic area. *Current Problems in Diagnostic Radiology*, 45, 80–83.
37. Marshet, E., Asamre, K., Bekele, J., Anteneh, T., Abera, M. (2011) The status of cystic echinococcosis (hydatidosis) in small ruminants slaughtered at Addis Ababa municipal abattoir. *Journal of Animal and Veterinary Advances*, 10, 1445–1449
38. Moon, S.L. and Khemalpure, S.S. (2017). Echinococcosis: current Indian scenario. *Global Journal of BioScience and Biotechnology*, 6, 383–389.
39. Moro, P. and Schantz, P.M. (2009) Echinococcosis: a review. *International Journal of Infectious Diseases*, 13, 125-133.
40. NCDC. (2016). Zoonotic Diseases of Public Health Importance. GOI. Accessed from [ncdc.gov.in](http://ncdc.gov.in) on 24<sup>th</sup> May 2020.
41. Nikale, N. (2014) Prevalence of echinococcosis in dogs and humans. MVSc Maharashtra Animal and Fishery Sciences University, Nagpur, India.
42. Parija, S.C. and Shivaprakash, M.R. (2000). Recent trends in the diagnosis of cystic Echinococcosis. *Journal of International Medical Science Academics*, 13, 27-31.
43. Otero-Abad, B., and Torgerson, P.R. (2013). A Systematic Review of the Epidemiology of Echinococcosis in Domestic and Wild Animals. *PLoS Neglected Tropical Diseases*, 7(6), e2249.
44. Pednekar, R.P., Gatne, M.L., Thompson, R.C., Traub, R.J. (2009). Molecular and morphological characterisation of *Echinococcus* from food producing animals in India. *Veterinary Parasitology*, 165, 58–65.
45. Pillai, K. J., Naryana Rao, P. L. and Surya Rao, K. (1986). A study on prevalence of hydatidosis in sheep and goats at Tirupati Municipal Slaughter house. *Indian Journal of Public Health*, 30, 160-165.
46. Prasad, B.N. (1981). Note on the pathology of hydatidosis among pigs in Bihar and its public health importance. *Haryana Veterinarian*, 20, 24-28.
47. Prathiush, P. R., D'Souza, P. E. and Gowda, A. K. J. (2008). Diagnosis of *E. granulosus* infection in dogs by a coproantigen sandwich ELISA. *Veterinarski Arhiv*, 78, 297-305.
48. Rana, U. V. S., Sehgal, S., Bhatia, R. and Bhardwaj, M. (1986). Hydatidosis in animals in and around Delhi. *Journal of Communicable Diseases*, 18, 116-119.
49. Rao, S.S., Mehra, B. and Narang, R. (2012). The spectrum of hydatid disease in rural central India: An 11-year experience. *Annals of Tropical Medicine and Public Health*, 5, 225-230.
50. Reddy, C.R.R.M., Morasiah, I.L., Parthavi, G. and Somasundara Rao, M. (1958). Epidemiology of hydatid disease in Kurnool. *Indian Journal of Medical Research*, 56, 1205-1220.
51. Rojas, C. A. A., Mathis, A., and Deplazes, P. (2018). Assessing the contamination of food and the environment with *Taenia* and *Echinococcus* eggs and their zoonotic transmission. *Current Clinical Microbiology Reports*, 5, 154-163.
52. Sajjan, S. A., Paturkar, A. M., Zende, R. J., Vaidya, V. M. and Chethan Kumar, H. B. (2015). Prevalence of hydatidosis in domestic pigs and elucidation of its economic impact on pork production. *Indian Veterinary Journal*, 92, 15-18.
53. Sangaran, A., Arunkumar, S. and John, L. (2014). Incidence of hydatidosis in slaughtered sheep and goats. *Indian Journal of Veterinary and Animal Science Research*, 43, 156-158.
54. Sarma, M. Dev, Deka, D. K. and Borkakoty, M. R. (2000) Occurrence of hydatidosis and porcine cysticercosis in Guwahati city. *Journal of Veterinary Parasitology*, 14, 173-174.
55. Shanmugam, S., Shanmugam, A. M. and Jayrajan, S. (1994). Incidence of morbid conditions of visceral organs of meat animals slaughtered at Muttur Dam Town. *International Journal of Animal Science*, 9, 101-103.
56. Sharma, M., Sehgal, R., Fomda, B.A., Malhotra, A. and Malla, N. (2013). Molecular characterization of *Echinococcus granulosus* cysts in North Indian Patients: Identification of G1, G3, G5 and G6 Genotypes. *PLoS Neglected Tropical Diseases*, 7, e2262.

57. Sharma, R., Sharma, D.K., Juyal, P.D., Aulakh, G.S. and Sharma, J.K. (2004). Pig Hydatidosis in and around Ludhiana City of Punjab. *Journal of Veterinary Public Health*, 2, 11-13.
58. Shekhawat, S. S., Purohit, S. K., Rao, R., Joshi, R. and Chaubey, H. (2005). Prevalence of hydatidosis in sheep and goats in Rajasthan. *Journal of Veterinary Public Health*, 3, 83-85.
59. Siles-Lucas, M., Casulli, A., Conraths, F. J. and Muller, N. (2017). Laboratory diagnosis of *Echinococcus* spp. in human patients and infected animals. In: *Advances in parasitology* 96: 159-257.
60. Singh, B.B., Dhand, N.K., Ghatak, S., Gill, J.P.S. (2014a). Economic losses due to cystic Echinococcosis in India: need for urgent action to control the disease. *Preventive Veterinary Medicine*, 113, 1–12.
61. Singh, B.B., Sharma, J.K., Ghatak, S., Sharma, R., Bal, M.S., Tuli, A., Gill, J.P.S. (2012). Molecular epidemiology of Echinococcosis from food producing animals in north India. *Veterinary Parasitology*, 186, 503–506.
62. Singh, B.B., Sharma, J.K., Tuli, A., Sharma, R., Bal, M.S., Aulakh, R.S., Gill, J.P.S. (2014b). Prevalence and morphological characterization of *Echinococcus granulosus* from north India. *Journal of Parasitic Diseases*, 38, 36–40.
63. Singh, B.P. and Dhar, D.N. (1988). *Echinococcus granulosus* in animals in Northern India. *Veterinary Parasitology*, 28, 261 – 266.
64. Socialcops (<https://blog.socialcops.com/intelligence/stray-dogs-in-india/>). Last accessed on 24 May, 2020.
65. Statista.com (<https://statista.com/statistics/india-population-of-pet-dogs/>). Last accessed on 24 May, 2020.
66. Sunita, T., Khurana, S., Malla, N. and Dubey, M.L. (2011). Immunodiagnosis of cystic echinococcosis by antigen detection in serum, urine, and saliva samples. *Tropical Parasitology*, 1, 33-38.
67. Thompson, R.C.A. and Lymbery, A.J. (1990). *Echinococcus*: Biology and strain variation. *International Journal of Parasitology*, 20, 457-470.
68. Thrusfield, M. (2007). *Veterinary Epidemiology* Ed. 3<sup>rd</sup> Blackwell Science Ltd. Ames, Iowa, USA.
69. Vaidya, V. M., Zende, R. J., Paturkar, A. M., Kumar, A. and Raut, C.K. (2014). Economic impact of hydatidosis and cysticercosis in food animals slaughtered at different abattoirs of Maharashtra. *Journal of Veterinary Public Health*, 12, 65-70.
70. Varma, T. K. (1990). Prevalence of *Echinococcus granulosus* infection in domestic animals of district Gurgaon (Haryana), India. *Philippine Journal of Veterinary Medicine*, 27, 65-69.
71. Varma, T. K. and Malviya, H.C. (1992). Studies on the fertility rate of hydatid cyst from domestic animals and prevalence of *Echinococcus granulosus* infestation amongst stray dogs. *Indian Journal of Parasitology*, 16, 55-57.
72. Vijayasmitha, R., Jagannath, M. S., Abdul Rahman, S. and Honnappa, T. G. (1993). The utility of leukocyte migration inhibition test in the diagnosis of hydatidosis in food animals. *Indian Journal of Animal Sciences*, 63, 596-599.
73. WHO, (2018). <http://www.who.int/news-room/fact-sheets/detail/echinococcosis>. Last accessed on September, 2019.
74. Zaman, K., Mewara, A., Kumar, S., Goyal, K., Khurana, S., Tripathi, P. and Sehgal, R. (2017). Seroprevalence of human cystic echinococcosis from North India (2004–2015). *Tropical Parasitology*. 7, 103–106.

\*\*\*\*\*